

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Lung adenocarcinoma

NAME Not Given

DATE OF BIRTH Not Given

SEX Not Given

MEDICAL RECORD # Not Given

PHYSICIAN

ORDERING PHYSICIAN Not Given MEDICAL FACILITY Not Given

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SPECIMEN

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Genomic Signatures

Microsatellite status - MS-Stable

Tumor Mutational Burden - TMB-Low (4 Muts/Mb)

Gene Alterations

For a complete list of the genes assayed, please refer to the Appendix.

ALK EML4-ALK fusion (Variant 1)

CCND1 amplification

FGF19 amplification

FGF3 amplification

FGF4 amplification

NFKBIA amplification

NKX2-1 amplification

TP53 R306*

7 Disease-relevant genes with no reportable alterations: EGFR, KRAS, BRAF, MET, RET, ERBB2, ROS1

6 Therapies Approved in the EU

14 Clinical Trials

O Therapies with Lack of Response

GENOMIC SIGNATURES

Microsatellite status - MS-Stable

Tumor Mutational Burden -

TMB-Low (4 Muts/Mb)

GENE ALTERATIONS

ALK - EML4-ALK fusion (Variant 1)

10 Trials see p. 11

CCND1 - amplification

4 Trials see p. 14

ACTIONABILITY

No therapies or clinical trials. see Genomic Signatures section

No therapies or clinical trials. see Genomic Signatures section

THERAPIES APPROVED IN THE EU THERAPIES APPROVED IN THE EU (IN PATIENT'S TUMOR TYPE) (IN OTHER TUMOR TYPE) Alectinib None Ceritinib Crizotinib None

FOUNDATIONONE®CDx

GENE ALTERATIONS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Gene Alterations section.

FGF19 - amplificationp. 5	NFKBIA - amplification	р. 6
FGF3 - amplificationp.5	NKX2-1 - amplification	. p. 6
FGF4 - amplificationp. 6	TP53 - R306*	p. 7

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved through a centralized EU procedure or a national procedure in an EU Member State. Therapies, including but not limited to the following, have been approved nationally and may not be available in all EU Member States: Tretinoin, Anastrozole, Bicalutamide, Cyproterone, Exemestane, Flutamide, Goserelin, Letrozole, Leuprorelin, Triptorelin.





GENOMIC SIGNATURES

GENOMIC SIGNATURE

Microsatellite status

CATEGORY MS-Stable

POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence, microsatellite stable (MSS) tumors are significantly less likely than MSI-high (MSI-H) tumors to respond to anti-PD-1 immune checkpoint inhibitors1-3, including approved therapies nivolumab and pembrolizumab4-5. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%,

p=0.001)6. Pembrolizumab therapy resulted in a significantly lower objective response rate (ORR) in MSS colorectal cancer (CRC) compared with MSI-H CRC (0% vs. 40%)5. Similarly, a clinical study of nivolumab, alone or in combination with ipilimumab, in patients with CRC reported a significantly higher response rate in patients with MSI-H tumors than those without4.

FREQUENCY & PROGNOSIS

MSI-high (MSI-H) has been reported at various frequencies in non-small cell lung cancer (NSCLC) as well as in small cell lung cancer7-12. One study observed MSI-H in 0.8% (4/480) of lung adenocarcinoma cases; the MSI-H tumors occurred in patients with smoking history, and 3 of the 4 MSI-H cases had nonsynchronous carcinomas in other organs, although none of the patients were diagnosed with Lynch syndrome7.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor13. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS213-15. The tumor seen here is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers16-18. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins13,15,17-18

GENOMIC SIGNATURES

GENOMIC SIGNATURE

Tumor Mutational Burden

CATEGORY
TMB-Low (4 Muts/Mb)

POTENTIAL TREATMENT STRATEGIES

On the basis of emerging clinical evidence, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-CTLA-419, anti-PD-L120-23, and anti-PD-1 therapies5,24-25; FDA-approved agents include ipilimumab, atezolizumab, avelumab, durvalumab, pembrolizumab, and nivolumab. In multiple solid tumor types, higher mutational burden has corresponded with response and improved prognosis. Pembrolizumab improved progression-free survival (14.5 vs. 3.4-3.7 months) for patients with non-small cell lung cancer (NSCLC) and higher mutational load (greater than 200 nonsynonymous mutations; hazard ratio = 0.19)25. In studies of patients with either NSCLC or colorectal cancer (CRC), patients whose tumors harbored elevated mutational burden reported higher overall response rates to pembrolizumab5,24-25. Anti-PD-1 therapies have achieved clinical benefit for certain patients with high mutational burden, including 3 patients with endometrial adenocarcinoma who reported sustained partial responses (PRs) following treatment with pembrolizumab26 or nivolumab27, a patient with hypermutant glioblastoma who obtained clinical benefit from pembrolizumab28, 2 pediatric patients with

biallelic mismatch repair deficiency-associated ultrahypermutant glioblastoma who experienced clinically and radiologically significant responses to nivolumab29, and 2 patients with microsatellite-stable rectal cancers, 1 who achieved an ongoing PR to pembrolizumab and the other an ongoing complete response to nivolumab30. For patients with melanoma, mutational load was associated with long-term clinical benefit from ipilimumab19,31 and anti-PD-1/anti-PD-L1 treatments21. For patients with metastatic urothelial carcinoma, those who responded to atezolizumab treatment had a significantly increased mutational load (12.4 mutations [muts] per megabase [Mb]) compared to nonresponders (6.4 muts/Mb)20, and mutational load of 16 muts/Mb or higher was associated with significantly longer overall survival22. In a retrospective analysis of 17 solid tumor types (comprised of 47% NSCLC, 40% mUC, and 13% encompassing 15 other solid tumors), a TMB of ≥16 muts/Mb associated with an objective response rate to atezolizumab of 30% vs. 14% for chemotherapy alone32.

FREQUENCY & PROGNOSIS

Intermediate TMB has been reported in 30-31% of non-small cell lung carcinomas (NSCLC), including 30% of adenocarcinomas and 41% of squamous cell carcinomas (SCC) (Spigel et al., 2016; ASCO Abstract 9017). Intermediate TMB was frequently observed in NSCLC with BRAF (31%) or KRAS (39%) mutation (Spigel et al., 2016; ASCO Abstract 9017). Although some studies have reported a lack of association between smoking and mutational burden in NSCLC (Schwartz et al., 2016; ASCO Abstract 8533)66,67, several other large studies did find a strong association with increased

Low TMB is observed more commonly in nonsmall cell lung carcinomas (NSCLC) harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are observed in approximately half of intermediate-high TMB cases33. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC34-36, several other large studies did find a strong association with increased TMB37-40. A large study of Chinese patients with lung adenocarcinoma reported a shorter median overall survival (OS) for tumors with a higher number of mutations in a limited gene set compared with lower mutation number (48.4 vs. 61.0 months)35.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma41-42 and cigarette smoke in lung cancer25,43, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes44-48, and microsatellite instability (MSI)44,47-48. The tumor seen here harbors a low TMB. Compared to patients with tumors harboring higher TMB levels, patients with tumors harboring low TMB levels have experienced lower rates of clinical benefit from treatment with immune checkpoint inhibitors, including anti-CTLA-4 therapy in melanoma19, anti-PD-L1 therapy in urothelial carcinoma20, and anti-PD-1 therapy in non-small cell lung cancer and colorectal cancer5,25.



GENE ALTERATIONS

GENE ALK

ALTERATION
EML4-ALK fusion (Variant 1)

POTENTIAL TREATMENT STRATEGIES

The ALK inhibitors crizotinib, ceritinib, brigatinib, and alectinib have shown significant clinical activity for patients with non-small cell lung cancer (NSCLC) whose tumors test positive for ALK rearrangement49-50 51-56. As first-line treatment, crizotinib improved overall survival (OS) relative to chemotherapy (HR=0.35) for patients with ALK+ advanced NSCLC57. Crizotinib has also shown activity in ALKmutant neuroblastoma58-59. Preclinically, ALK activating point mutations are crizotinibsensitive60-61. A Phase 1 study of ceritinib in ALK-rearranged NSCLC reported overall survival (OS) of 72% (60/83) for patients who were ALK inhibitor-naive and median progression-free survival (PFS) of 18.4 months, versus an OS of 56% (92/163) and PFS of 6.9 months for those who were previously treated62. A Phase 1/2 study of brigatinib for patients with ALK-rearranged NSCLC reported confirmed ORRs of 62% (44/71) and 100% (8/8) for crizotinib-treated and crizotinib-naive patients, respectively53. Antitumor activity was also seen in the central nervous system (CNS), a common site of failure during crizotinib treatment53,63-64. Alectinib combined with atezolizumab led to an ORR of 81% (17/21) as first-line treatment for PD-L1 unselected, ALK+ NSCLC65. Lorlatinib led to an ORR of 73% (43/59), 39% (11/28), and 39% (43/111), and intracranial ORR of 68% (25/37), 46% (6/13), and 47% (38/81), for patients with NSCLC previously treated with crizotinib, one

prior ALK inhibitor, or 2-3 prior ALK inhibitors, respectively66. For patients whose tumors harbored one or more ALK kinase domain mutations, lorlatinib led to responses for 64% (29/45), including 58% (11/19) for those with the ALK G1202R resistance mutation67; G1202 therefore does not appear to represent a major mechanism of lorlatinib resistance68-69. Lorlatinib led to complete resolution of intrathecal metastases and stabilization of CNS metastases for a heavily pretreated patient with ALK+ NSCLC70, and its use in the fourth-line setting led to disappearance of leptomeningeal disease for a patient with ALK-rearranged metastatic inflammatory myofibroblastic sarcoma71. The combination of lorlatinib and the PD-L1 inhibitor avelumab led to a confirmed response rate of 46.4% [12 partial responses (PRs), 1 complete response] for the 28 patients with ALK+ NSCLC who were treated72. Ensartinib treatment for ALK+ NSCLC led to ORRs of 80%, 69%, and 64% for patients who were treatment-naive, crizotinib refractory, or for intracranial metastases, respectively73. Phase 1 studies of the ALK/ROS1/TRK inhibitor entrectinib have reported responses for 4/7 (57%) kinase inhibitor-naive patients with ALK-rearranged solid tumors, including patients with NSCLC, renal cell carcinoma, and colorectal cancer; as well as for 1 patient with ALK F1245V mutant neuroblastoma but in o/13 patients with ALK fusion-positive tumors previously treated with an ALK inhibitor and in none of the other patients with ALK non-fusion alterations 74. A Phase 2 trial of the HSP90 inhibitor ganetespib reported PRs in a small number of patients with ALK-rearranged NSCLC75.

FREQUENCY & PROGNOSIS

The EML4-ALK gene fusion has been observed in approximately 3-7% of non-small cell lung cancer (NSCLC) cases, more frequently in vounger patients. non-smokers. males. and

patients of Asian heritage 76-82. Other rearrangements involving ALK have also been described in lung cancer 83-84. EML4-ALK fusions have been reported to be a significant indicator of poor prognosis in advanced stage NSCLC 82.

FINDING SUMMARY

ALK encodes a receptor tyrosine kinase, a member of the insulin receptor superfamily, whose activation induces the downstream pathways associated with cell survival, angiogenesis, and cell proliferation85. Different EML4-ALK variants have been identified in cancer, all of which contain the intracellular tyrosine kinase domain of ALK86. The most commonly observed rearrangements consist of ALK exon 20 fused to a variety of breakpoints in EML4: exon 13 (variant 1, 33-54% of cases)87-89, exon 20 (variant 2, 10-12% of cases)87-89, exon 6 (variant 3 a/b, 26-30% of cases)52,87-88,90, exon 15 (variant 4, 2% of cases)76,91-92, exon 18 (variant 5, 1.6-3% of cases)89,91, exon 2 (variant 5 a/b, 1-2% of cases)87,92-94, and exon 17 (variant 8 a/b, <1%)89,91,95. All of these variants have been characterized as, or are predicted to be, activating and sensitive to ALK inhibitors, including crizotinib and ceritinib88,90,96; however, variants 3a/b are less sensitive to crizotinib in vitro88. Although EML4-ALK variant 1 was associated with significantly longer median progression-free survival (11 months vs. 4.2 months) in a small study of crizotinib-treated non-small cell lung cancer (NSCLC)97, other studies have not found a correlation between EML4-ALK variants and response to crizotinib in NSCLC52,89.

GENE ALTERATIONS

CCND1

amplification

POTENTIAL TREATMENT STRATEGIES

Amplification or overexpression of CCND1 may predict sensitivity to CDK4/6 inhibitors, such as FDA-approved abemaciclib, palbociclib,

and ribociclib98-99 100-101102-105. Clinical benefit has been reported for patients with solid tumors with CCND1 amplification or expression in response to treatment with palbociclib106, ribociclib98-99 100,104, and abemaciclib105.

FREQUENCY & PROGNOSIS

In the TCGA dataset, amplification of CCND1 has been found in 4.3% of lung adenocarcinoma cases107. Other studies have reported CCND1 amplification in 3-25% of lung adenocarcinomas108-109. Expression of cyclin D1 has been reported in 59% (36/61) of

non-small cell lung cancer tumors analyzed but was not reported to be associated with clinicopathologic parameters110.

FINDING SUMMARY

CCND1 encodes cyclin D1, a binding partner of the kinases CDK4 and CDK6, that regulates RB activity and cell cycle progression.

Amplification of CCND1 has been positively correlated with cyclin D1 overexpression111 and may lead to excessive proliferation112-113.

FGF19

amplification

POTENTIAL TREATMENT STRATEGIES

There are no targeted therapies that directly address genomic alterations in FGF19. However, amplification of FGF19 predicts sensitivity to inhibitors of FGF19 predicts sensitivity to inhibitors of FGFR4 in liver cancer cell lines114; in one preclinical study, selective inhibition of FGFR4 reduced tumor burden in an FGF19-amplified HCC xenograft model115. A Phase 1 study of the FGFR4 inhibitor BLU-554 for previously treated HCC (11/14 sorafenib) reported 1 partial response and 1 stable disease (SD) in patients with FGF19-positive HCC116. Preliminary results from the dose escalation part of a Phase 1/2 study evaluating another FGFR4 inhibitor,

FGF401, showed an overall response rate of 8% (4/53), 53% (28/53) SDs, and a median time to progression of 4.1 months; responses were observed in both FGF19-positive and -negative cases117. In one clinical study, a trend toward response to sorafenib treatment and FGF19 copy number gain was observed in patients with HCC, and 2 patients harboring FGF19 copy number gain experienced a complete response118. Multiple therapies targeting FGF19 or FGFR4 signaling are in preclinical development119, and clinical trials evaluating inhibitors of FGFR4 are under way for patients with solid tumors.

FREQUENCY & PROGNOSIS

In the TCGA datasets, FGF19 amplification has been reported with highest incidence in esophageal carcinoma (35%), head and neck squamous cell carcinoma (28%), breast carcinoma (16%), lung squamous cell carcinoma (12%), bladder urothelial carcinoma (12%), and cholangiocarcinoma (11%) (cBioPortal, 2017). In HCC, FGF19 is an

important driver gene115,120-121, and FGF19 protein expression correlates with tumor progression and poorer prognosis122. Exogenous FGF19 has been shown to promote prostate cancer tumorigenesis in a preclinical study123, and the presence of FGF19-positive tissues is an independent factor for worse prognosis following radical prostatectomy124.

FINDING SUMMARY

FGF19 encodes fibroblast growth factor 19, an FGFR4 ligand involved with bile acid synthesis and hepatocyte proliferation in the liver115,125. FGF19 lies in a region of chromosome 11q13 frequently amplified in a diverse range of malignancies that also contains FGF3, FGF4, and CCND1126. Correlation between FGF19 amplification and protein expression has been demonstrated in hepatocellular carcinoma (HCC)127 but was not observed in several other tumor types120.

FGF3

amplification

POTENTIAL TREATMENT STRATEGIES

There are no targeted therapies that directly address genomic alterations in FGF3. Inhibitors of FGF receptors, however, are

undergoing clinical trials in a number of different cancers.

FREQUENCY & PROGNOSIS

FGF3 lies in a region of chromosome 11q13 that also contains FGF19, FGF4, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression. This chromosomal region is frequently amplified in a diverse range of malignancies112.

FINDING SUMMARY

FGF3 encodes fibroblast growth factor 3, a growth factor that plays a central role in

development of the inner ear. Germline mutations in FGF3 give rise to an autosomal recessive syndrome characterized by microdontia, deafness, and complete lack of inner ear structures128.

GENE ALTERATIONS

FGF4

amplification

POTENTIAL TREATMENT STRATEGIES

FGF4 amplification and overexpression was associated with cell sensitivity to the multikinase inhibitor sorafenib in preclinical studies129-130 and amplification of FGF4/FGF3 in HCC significantly correlated with patient response to sorafenib (p=0.006)129. Therefore, thyroid carcinoma. Sorafenib is under

investigation in clinical trials in multiple tumor types. FGF4 amplification may confer sensitivity to sorafenib, which is FDA approved to treat HCC, renal cell carcinoma, and differentiated

FREQUENCY & PROGNOSIS

This chromosomal region is frequently amplified in a diverse range of malignancies112 including esophageal carcinoma (35%), head and neck squamous cell carcinoma (HNSCC; 28%), breast invasive carcinoma (16%), lung squamous cell carcinoma (12%), bladder urothelial carcinoma (12%), ovarian serous cystadenocarcinoma (8%), stomach adenocarcinoma (7%), skin melanoma (6%), and hepatocellular carcinoma (HCC; 5%) (cBioPortal, 2017).

FINDING SUMMARY

FGF4 encodes fibroblast growth factor 4, which plays a central role in development of the teeth131 and acts synergistically with other FGFs and SHH (sonic hedgehog) to regulate limb outgrowth in vertebrate development132. FGF4 lies in a region of chromosome 11q13 that also contains FGF19, FGF3, and CCND1, the latter gene encoding cyclin D1, a key regulator of cell cycle progression.

Amplification of FGF4, along with that of FGF3, FGF19, and CCND1, has been reported in a variety of cancers112,129,133-136 and may confer sensitivity to the multi-kinase inhibitor sorafenib129.

NFKBIA

amplification

POTENTIAL TREATMENT STRATEGIES

There are no therapies that directly target NFKBIA amplification or expression.

FREQUENCY & PROGNOSIS

In the TCGA datasets, amplification of NFKBIA has been reported with the highest incidence in lung adenocarcinoma (11.7%)107, esophageal carcinoma (3.8%), uterine carcinosarcoma (3.6%), lung squamous cell carcinoma (3.4%), and ovarian serous cystadenocarcinoma (2.6%) (cBioPortal, 2017). Amplification or increased expression of NFKBIA in EGFR-mutant lung cancer has been reported to predict improved response to EGFR tyrosine kinase inhibitors137-138. Certain NFKBIA polymorphisms, which may affect lkBa expression levels, have been studied as

risk factors for some cancer types, although the data are mixed and conflicting 130-141.

FINDING SUMMARY

NFKBIA encodes IkBa, an inhibitor of the NF-kappaB (NFkB)/REL complex. It has been reported to act as a tumor suppressor in Hodgkin's lymphoma 395-399 and in glioblastoma 392,400-401. NFKBIA has been reported to be amplified in cancer 227 and may be biologically relevant in this context 228-229. In contrast, truncating mutations that result in loss of the majority of the IkBa protein are predicted to be inactivating.

NKX2-1

amplification

POTENTIAL TREATMENT STRATEGIES

There are no approved therapies or trials that target tumors with TTF-1 amplification or overexpression. Lung cancer cell lines that express both TTF-1 and NKX2-8, which is located in the same amplicon as NKX2-1, have demonstrated resistance to cisplatin therapy152,

although conflicting data has also been reported153.

FREQUENCY & PROGNOSIS

Putative amplification of NKX2-1 has been reported with the highest incidence in lung cancer, and has been observed in 14% of adenocarcinomas107 and 5% of squamous cell carcinomas (SCC)154 as well as other tumor types including prostate adenocarcinomas (6%)155, and poorly differentiated and anaplastic thyroid cancers (4%)156. NKX2-1 mutation has been observed in 9% of acinar cell carcinomas of the pancreas157, 5% of uterine carcinosarcomas158, and is infrequent in other tumor types (cBioPortal, COSMIC, 2018). TTF-1 is expressed in a majority of lung adenocarcinomas and small cell carcinomas, as

well as in a subset of thyroid and CNS tumors159-161. Cytoplasmic TTF-1 expression has been reported as an adverse prognostic factor in breast carcinoma162-163. However, whether amplification and/or expression status of NKX2-1 have prognostic implications for patients with lung cancer is controversial152-153,164-167. TTF-1 has been observed to have tumor-promoting as well as anti-oncogenic roles168-169.

FINDING SUMMARY

NKX2-1 (NK2 homeobox 1) encodes the thyroid transcription factor TTF-1170. Amplification of NKX2-1 results in overexpression of TTF-1 and upregulated transcription of downstream target genes171.

GENE ALTERATIONS

TP53

ALTERATION R306*

POTENTIAL TREATMENT STRATEGIES

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor AZD1775172-175 or p53 gene therapy and immunotherapeutics such as SGT-53176-180 and ALT-801181. In a Phase 1 study, AZD1775 in combination with gemcitabine, cisplatin, or carboplatin elicited partial response in 10% (17/176) and stable disease in 53% (94/176) of patients with solid tumors; the response rate was 21% (4/19) in patients with TP53 mutations versus 12% (4/ 33) in patients who were TP53-wild-type182. Combination of AZD1775 with paclitaxel and carboplatin achieved significantly longer progression-free survival than paclitaxel and carboplatin alone in patients with TP53-mutant ovarian cancer183. Furthermore,

AZD1775 in combination with carboplatin achieved a 27% (6/22) response rate and 41% (9/22) stable disease rate in patients with TP53-mutant ovarian cancer refractory or resistant to carboplatin plus paclitaxel184. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including two confirmed and one unconfirmed partial responses and two instances of stable disease with significant tumor shrinkage180. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53 mutant, but not TP53 wild-type, breast cancer xenotransplant mouse model185. Clinical trials of these agents are under way for some tumor types for patients with a TP53 mutation.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer. TP53 mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)107,154,186-191. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma192. In one study of 55 patients with lung adenocarcinoma, TP53 alterations

correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors pembrolizumab and nivolumab in this study24.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers193. Any alteration that results in the disruption or partial or complete loss of the region encoding the TP53 DNA-binding domain (DBD, aa 100-292) or the tetramerization domain (aa 325-356), such as observed here, is thought to dysregulate the transactivation of p53-dependent genes and is predicted to promote tumorigenesis194-196. Germline mutations in TP53 are associated with the very rare disorder Li-Fraumeni syndrome and the early onset of many cancers197-202. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000203 to 1:20,000202, and in the appropriate clinical context, germline testing of TP53 is recommended.



THERAPIES APPROVED IN THE EU

IN PATIENT'S TUMOR TYPE

Alectinib

Assay findings associations

ALK

EML4-ALK fusion (Variant 1)

AREAS OF THERAPEUTIC USE

Alectinib is a tyrosine kinase inhibitor that targets ALK and RET. It is available in the EU to treat patients with ALKpositive advanced non-small cell lung cancer (NSCLC) as first-line therapy or after prior treatment with crizotinib.

GENE ASSOCIATION

Activating ALK alterations may predict sensitivity to alectinib on the basis of extensive clinical evidence in ALK-rearranged NSCLC51,20456,205-206.

SUPPORTING DATA

Alectinib has been primarily studied for the treatment of ALK-rearranged NSCLC. In the Phase 3 ALEX study comparing alectinib with crizotinib in ALK-rearranged, inhibitor-naive NSCLC, patients treated with alectinib experienced significantly improved progression-free survival (PFS), 68.4% versus 48.7% (hazard ratio [HR]=0.47); median PFS was not reached in the alectinib arm and was 11.1 months in the crizotinib arm; and median overall survival (OS) was not reached in either arm at 2 years207. Similar results have been reported in the J-ALEX trial for inhibitor-naive Japanese patients with ALK-positive NSCLC208. Alectinib combined with atezolizumab led to an objective response rate (ORR) of 81% (17/21) as first-line treatment for PD-L1 unselected, ALK+ NSCLC65. In the context of crizotinib resistance, the Phase 3 ALUR trial for patients with ALK+ NSCLC

reported that alectinib significantly improved PFS relative to chemotherapy (7.1 vs. 1.6 months; HR=0.32)209. Phase 1/2 and Phase 2 trials of alectinib in ALK-rearranged NSCLC refractory to crizotinib reported ORRs of 45-55%56,206,210, with a reported median duration of response of 11.2-17 months56,210-211. Alectinib has demonstrated significant activity against central nervous system (CNS) metastases, such as leptomeningeal metastases, for patients with NSCLC56,204-207,210,212-216. In the ALUR trial, alectinib significantly improved ORR for CNS metastases relative to chemotherapy (54.2% vs. 0%)209. In the ALEX study, alectinib showed superior efficacy in CNS compared with crizotinib, with 12-month progression rate with CNS disease of 41.4% versus 9.4% and median duration of response in patients with CNS disease at baseline for 17.3 months versus 5.5 months207. A Phase 2 study of alectinib for crizotinib-refractory, ALK rearranged NSCLC reported 27% of patients achieving a CNS-specific CR, and an overall CNS disease control rate of 83% (95% confidence interval, 74% to 91%)56. In a preliminary study of alectinib in four cases of metastatic, RET-rearranged NSCLC, three of whom had previously been treated with cabozantinib, PRs were observed in two patients (one confirmed and one unconfirmed), with an additional patient exhibiting SD for 6 weeks and one case of progressive disease; improvement in CNS disease was observed in one patient after dose increase217.

Ceritinib

Assay findings associations

ALK

EML4-ALK fusion (Variant 1)

AREAS OF THERAPEUTIC USE

Ceritinib is an inhibitor of the kinases ALK, ROS1, IR, and IGF-1R. It is available in the EU to treat advanced ALKpositive non-small cell lung carcinoma (NSCLC) either as first-line treatment or following crizotinib therapy.

GENE ASSOCIATION

On the basis of strong clinical data demonstrating benefit to patients with crizotinib-naïve lung cancer62,223-224 or those previously treated with crizotinib225-22655,62,227, ALK rearrangements may predict sensitivity to ceritinib.

SUPPORTING DATA

Multiple Phase 3 studies have reported clinical benefit from ceritinib for patients with advanced ALK-rearranged (ALK+) NSCLC. As a first-line treatment for patients with ALK+ NSCLC in the ASCEND-4 Phase 3 study, ceritinib monotherapy significantly increased the median progression-free survival (PFS) to 16.6 months, compared to a median PFS of 8.1 months in patients with platinumbased chemotherapy224. A Phase 3 study of ceritinib for ALK inhibitor-naïve patients with ALK+

NSCLC observed a whole-body (WB) objective response rate (ORR) of 63.7%, a WB disease control rate (DCR) of 89.5%, and progression-free survival (PFS) of 11.1 months223. The ASCEND-5 Phase 3 study comparing ceritinib to chemotherapy for patients with ALK+ NSCLC previously treated with crizotinib and chemotherapy also reported a significant benefit for ceritinib in ORR (39% vs. 7%) and median PFS (5.4 vs. 1.6 months); there was no improvement of median OS (18.1 vs. 20.1 months), which may be due to the crossover of patients to the ceritinib arm226. The ASCEND-1 Phase 1 study of ceritinib for patients with ALK+ NSCLC reported an ORR of 72%, median PFS of 18.4 months, and 12-month overall survival (OS) of 83%62. Earlier Phase 1 and 2 studies reported similar clinical benefit as measured by ORR (39-57%), median PFS (5.7-6.9 months), and median OS of 16.7 months55,62,227; for patients with brain metastases, an intracranial ORR of 39% and duration of response of 12.8 months were achieved225. Case studies have also reported responses to ceritinib in patients with ALK+ NSCLC and ALK missense mutation after disease progression on crizotinib228 or alectinib229-230.



THERAPIES APPROVED IN THE EU

IN PATIENT'S TUMOR TYPE

Crizotinib

Assay findings associations

ALK

EML4-ALK fusion (Variant 1)

AREAS OF THERAPEUTIC USE

Crizotinib is an inhibitor of the kinases MET, ALK, ROS1, and RON. It is available in the EU to treat patients with advanced non-small cell lung cancer (NSCLC) whose tumors are positive for ALK either as first-line or following previous treatment. It is also available to treat patients with ROS1-positive advanced NSCLC.

GENE ASSOCIATION

ALK activation may predict sensitivity to crizotinib. In patients with ALK-rearranged NSCLC, crizotinib improved outcomes in both the first-line231-232 and second-line54 settings compared with chemotherapy. Retrospective analysis of 35 patients with NSCLC indicated that compared with other EML4-ALK variants, EML4-ALK variant 1 was an independent predictor of improved median PFS (11.0 vs. 4.2 months, hazard ratio of 0.35) on crizotinib treatment97. ALK inhibitors have also demonstrated clinical activity in the context of several other cancer types with activating ALK alterations, including thyroid carcinoma, inflammatory myofibroblastic tumors, and anaplastic large cell lymphoma58,233-234.

SUPPORTING DATA

The Phase 3 PROFILE 1014 study for patients with ALK positive non-squamous NSCLC reported significantly prolonged progression-free survival [PFS, 10.9 vs. 7.0 months, hazard ratio (HR) 0.45] and higher objective response rate (ORR, 74% vs. 45%) with first-line crizotinib compared with pemetrexed and cisplatin or carboplatin232. A similar Phase 3 study for East Asian patients confirmed that crizotinib is superior to chemotherapy in this setting (PFS of 11.1 vs. 6.8 months, HR 0.40; ORR of 87.5% vs. 45.6%)231. In the ongoing

Phase 3 PROFILE 1007 study for patients with ALKpositive advanced NSCLC and prior platinum-based therapy (NCT00932893), crizotinib significantly improved median PFS (7.7 months vs. 3.0 months), ORR (65% vs. 20%), and quality of life as compared with chemotherapy54,235. The three Phase 3 studies observed numerical, but not statistically significant, improvement of overall survival (OS) with crizotinib (HR of o.82-o.90), although most patients (70-89%) crossed over from the chemotherapy groups to crizotinib treatment231,236232. The efficacy of crizotinib in patients with brain metastases has also been examined. Prospective comparison of the intracranial efficacy in patients with stable treated brain metastases included in PROFILE 1014 reported significantly prolonged intracranial disease control rate (DCR) at 24 weeks (56% vs. 25%) and PFS (9.0 vs. 4.0 months, HR 0.40) for patients treated with first-line crizotinib as compared with chemotherapy237. Pooled retrospective analysis of patients with ALK-rearranged NSCLC and concurrent brain metastases from the PROFILE 1007 and 1005 studies reported 12-week intracranial DCRs of 56% vs. 62% and intracranial ORR of 18% vs. 33% in patients with previously untreated versus previously treated brain metastases238. In a retrospective study of patients with brain metastases from ALK rearranged NSCLC, the majority of whom were treated with radiotherapy and crizotinib, the median OS after diagnosis of brain metastasis was 49.5 months; lack of prior targeted therapy, absence of extracranial metastasis, and a Karnofsky performance score of 90 or higher were significantly associated with improved OS239. Upon disease progression, further survival benefit can be derived for patients with ALK-positive NSCLC who continue crizotinib treatment240.





CLINICAL TRIALS

IMPORTANT Clinical trials are ordered by gene and prioritized in the following descending order: Pediatric trial qualification → Geographical proximity → Trial phase → Trial verification within last 2 months. While every effort is made to ensure the accuracy of the information

contained below, the information available in the public domain is continually updated and should be investigated by the physician or research staff. The clinical trials listed in this report may not be complete and exhaustive or may include trials for which the patient does not meet the

clinical trial enrollment criteria For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov or local registries in your region.

GENE ALK

ALTERATION
EML4-ALK fusion (Variant 1)

RATIONALE

ALK rearrangements, activating mutations, or amplification may be associated with increased activity in the ALK kinase. Therefore, drugs that inhibit ALK kinase may be relevant. Additionally, patients who have become resistant to crizotinib may harbor sensitivity to newer ALK inhibitors or to HSP90 inhibitors. Examples of clinical trials that may be appropriate for this patient are listed

below. These trials were identified through a search of the trial website clinicaltrials.gov using keyword terms such as "alectinib", "AF802", "CH5424802", "ceritinib", "LDK378", "crizotinib", "PF-02341066", "CEP-37440", "dalantercept", "gilteritinib", "ASP2215", "PF-06463922", "RXDX-101", "TSR-011", "X-396", "lung", "solid tumor", and/or "advanced cancer".

NCT03178552 PHASE 2 / 3

A Phase II/III Multicenter Study Evaluating the Efficacy and Safety of Multiple Targeted Therapies as Treatments for Patients With Advanced or Metastatic Non-Small Cell Lung Cancer (NSCLC) Harboring Actionable Somatic Mutations Detected in Blood (B-FAST: Blood-First Assay Screening Trial)

TARGETS
ALK, PD-L1, RET

LOCATIONS: Okayama (Japan), Shizuoka (Japan), Saga (Japan), Aichi (Japan), Hiroshima (Japan), Kurralta Park (Australia), Rio de Janeiro (Brazil), California, Krakow (Poland), Moscovskaya Oblast (Russian Federation), CD Mexico (Mexico), Kyoto (Japan), Malaga (Spain), Connecticut, San Luis Potosí (Mexico), Miyagi (Japan), Gdańsk (Poland), Santiago de Compostela (Spain), Warszawa (Poland), Madrid (Spain), Osaka (Japan), Esslingen (Germany), Bunkyo-ku (Japan), Ijui (Brazil), Ishikawa (Japan), Yamaguchi (Japan), Alicante (Spain), Barcelona (Spain), Shatin (Hong Kong), Hospitalet de Llobregat (Spain), Poitiers (France), Pennsylvania, Tokyo (Japan), Valencia (Spain), Toronto (Canada), Fukuoka (Japan), New York, Wakayama (Japan), Milano (Italy), Beer Sheva (Israel), Olsztyn (Poland), Florida, Illinois, Niigata (Japan), Ehime (Japan), Kanagawa (Japan), Otwock (Poland)

NCT02767804

Phase 3 Randomized Study Comparing X-396 to in Anaplastic Lymphoma
Kinase (ALK) Positive Non-Small Cell Lung Cancer (NSCLC) Patients

TARGETS
ABL, MET, ALK, ROS1, AXL, TRKC, TRKA

LOCATIONS: Pergamino (Argentina), Virginia, Changchun (China), Barcelona (Spain), Wisconsin, Nanchang (China), São Paulo (Brazil), Changsha (China), Bristol (United Kingdom), Santo André (Brazil), Jerusalem (Israel), Tianjin (China), New York, Warsaw (Poland), Rosario (Argentina), Oregon, Florida, Montpellier (France), Palma de Mallorca (Spain), Edirne (Turkey), Sondrio (Italy), Shenyang (China), Plesice (Czechia), Brussels (Belgium), Ostrava-Vitkovice (Czechia), Gdańsk (Poland), Hong Kong (Hong Kong), Haifa (Israel), Nottingham (United Kingdom), Qingdao (China), Moscow (Russian Federation), Hangzhou (China), Ravenna (Italy), Aviano (Italy), Missouri, Tennessee, Meldola (Italy), Nanjing (China), Idaho, Georgia, Hefei (China), Istanbul (Turkey), Legnago (Italy), Berlin (Germany), Usti nad Labem (Czechia), Beijing (China), Omsk (Russian Federation), Guangzhou (China), Buenos Aires (Argentina), Michigan, Milano (Italy), Lima (Peru), Saint Petersburg (Russian Federation), Pamplona (Spain), Madrid (Spain), Wuhan (China), Izmir (Turkey), Seoul (Korea, Republic of), Caba (Argentina)

NCT02568267

An Open-Label, Multicenter, Global Phase 2 Basket Study of for the Treatment of Patients With Locally Advanced or Metastatic Solid Tumors That Harbor NTRK1/2/3, ROS1, or ALK Gene Rearrangements

PHASE 2

TARGETS
ALK, ROS1, TRKC, TRKB, TRKA

Florida, Toulouse (France), Oklahoma, Kashiwa-shi (Japan), Washington, Lille (France), Michigan, Illinois, Gdansk (Poland), Barcelona (Spain), Ehime (Japan), Wisconsin, Georgia, Taipei (Taiwan), Köln (Germany), Albury (Australia), Maryland, Göttingen (Germany), Genova (Italy), Warszawa (Poland), Utah, North Carolina, Oregon, New Hampshire, Missouri, Padova (Italy), Madrid (Spain), Bedford Park (Australia), Gliwice (Poland), Tainan (Taiwan), Chang Hua (Taiwan), Hawaii, Amsterdam (Netherlands), Torino (Italy), Massachusetts, Orbassano (Italy), Roma (Italy), Arizona, Shatin (Hong Kong), Taichung (Taiwan), Villejuif cedex (France), Singapore (Singapore), Connecticut, Aichi (Japan), Marseille cedex 5 (France), Shizuoka (Japan), Otwock (Poland), Pisa (Italy), Poznań (Poland), Cheongju-si (Korea, Republic of), Candiolo (Italy), Nevada, Kowloon (Hong Kong), Bordeaux (France), Dresden (Germany),

LOCATIONS: Hyogo (Japan), London (United Kingdom), Lyon (France), Leiden (Netherlands), Taipei City (Taiwan), Cambridge (United Kingdom),

Virginia, Paris (France), Napoli (Italy), District of Columbia, Heidelberg (Australia), New Lambton Heights (Australia), Malaga (Spain), Montpellier cedex 5 (France), Berlin (Germany), Colorado, Paris cedex 15 (France), Miyagi (Japan), Texas, California, Liverpool (Australia), Manchester (United Kingdom), Ohio, Sevilla (Spain), Fukuoka (Japan), Osaka (Japan), Minnesota, Marseille (France), Fuenlabrada (Spain), Milano (Italy), Niigata (Japan), Perugia (Italy), New York, Seoul (Korea, Republic of), Hong Kong (Hong Kong)



CLINICAL TRIALS

NCT03093116	PHASE 1 / 2
A Phase 1/2, Open-Label, Multi-Center, First-in-Human Study of the Safety, Tolerability, Pharmacokinetics, and Anti-Tumor Activity of in Patients With Advanced Solid Tumors Harboring ALK, ROS1, or NTRK1-3 Rearrangements (TRIDENT-1)	TARGETS ALK, ROS1, TRKC, TRKB, TRKA

LOCATIONS: Massachusetts, Colorado, New York, Seoul (Korea, Republic of), California

NCT00585195	PHASE 1
Phase 1 Safety, Pharmacokinetic And Pharmacodynamic Study Of A C-met/Hgfr Selective Tyrosine Kinase Inhibitor, Administered Orally To Patients With Advanced Cancer	TARGETS MET, ALK, ROS1, AXL, TRKC, TRKA

LOCATIONS: New York, Michigan, Colorado, Ohio, Pennsylvania, California, Kashiwa (Japan), Nagoya (Japan), Akashi (Japan), Massachusetts, Melbourne (Australia), North Carolina, Seoul (Korea, Republic of), Vermont, Sapporo (Japan), Osakasayama (Japan)

NCT02693535	PHASE 2
Targeted Agent and Profiling Utilization Registry (TAPUR) Study	TARGETS ABL, CDK4, PARP, EGFR, DDR2, PDGFRs, VEGFRs, CTLA-4, ROS1, CSF1R, ERBB2, PD-1, ERBB3, MEK, RAF1, KIT, AXL, SMO, TRKC, mTOR, TRKA, MET, ALK, BRAF, RET, SRC, FLT3, CDK6

LOCATIONS: North Dakota, Washington, Illinois, California, Pennsylvania, Georgia, Arizona, Utah, North Carolina, Oklahoma, Alabama, South Dakota, Florida, Michigan, Oregon, Virginia, Texas, Nebraska

NCT01625234		PHASE 1 / 2
Phase 1/2, First-in-Human, Dose-Escalation Study of Solid Tumors and Expansion Phase in Patients With ALK-positive	in Patients With Advanced e Non-Small Cell Lung Cancer	TARGETS ABL, MET, ALK, ROS1, AXL

LOCATIONS: California, Oregon, Wisconsin, Tennessee, New York, Missouri, Maryland, South Carolina, Pennsylvania, Massachusetts, Texas, Virginia, Ohio, West Virginia

NCT02706626	PHASE 2
Phase 2 Trial of After Treatment With Second-Generation ALK Inhibitors in Refractory ALK Rearranged Non-Small Cell Lung Cancer (NSCLC)	TARGETS EGFR, ALK, ROS1
LOCATIONS: Colorado, Tennessee, Texas, North Carolina	

PHASE 1
TARGETS ALK, ROS1, mTOR



CLINICAL TRIALS

NCT02227940	PHASE 1
A Phase I Study of a Novel ALK Inhibitor, in Combination With Based Chemotherapy in Patients With Advanced Solid Tumors	TARGETS ALK, ROS1
LOCATIONS: New York	





CLINICAL TRIALS

GENE CCND1

ALTERATION amplification

RATIONALE

CCND1 amplification may activate CDK4/6 and may predict sensitivity to CDK4/6 inhibitors. Examples of clinical trials that may be appropriate for this patient are listed below. These trials were identified through a search of the trial website

clinicaltrials.gov using keyword terms such as "CDK4", "CDK6", "palbociclib", "PD-0332991", "abemaciclib", "LY2835219", "ribociclib", "LEE011", "NSCLC", "lung", "solid tumor", and/or "advanced cancer".

NCT03099174

An Open Label, Phase Ib Dose-escalation Study Evaluating the Safety and Tolerability of and Abemaciclib in Patients With Locally Advanced or Metastatic Solid Tumors and in Combination With Endocrine Therapy in Patients With Locally Advanced or Metastatic Hormone Receptorpositive Breast Cancer, Followed by Expansion Cohorts

TARGETS CDK4, Aromatase, ER, IGF-2, IGF-1, CDK6

LOCATIONS: Nevada, Madrid (Spain), Connecticut, Pozuelo de Alarcón (Spain), Paris (France), Marseille (France), Barcelona (Spain), California,

NCT02897375

A Phase 1 Study of in Combination With or in Advanced Solid

TARGETS CDK4, CDK6

PHASE 1

PHASE 1

LOCATIONS: Georgia

Malignancies

NCT01037790

Phase II Trial of the Cyclin-Dependent Kinase Inhibitor in Patients With Cancer

PHASE 2 **TARGETS**

CDK4, CDK6

LOCATIONS: Pennsylvania

NCT03065062

Phase I Study of the CDK4/6 Inhibitor in Combination With the PI3K/mTOR Inhibitor for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

PHASE 1 **TARGETS**

CDK4, mTORC1, PI3K-gamma, mTORC2, PI3K-alpha, CDK6

LOCATIONS: Massachusetts



APPENDIX

Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

 ASXL1
 CREBBP
 ERBB2
 FANCC

 N986S
 K2075R
 E503K
 C206F

KDM5CMEN1NOTCH1SMARCA4R1435CamplificationD1953HQ347K



APPENDIX

Genes assayed in FoundationOne $^{\otimes}$ CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	ATRX	AMER1 (FAM123B)
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRFI1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDM5A	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2	PARK2
PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD1)	PDCD1LG2 (PD-L2)	PDGFRA	PDGFRB
PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2	POLD1
POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1	PTEN
PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C	RAD51D
RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET	RICTOR
RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2	SF3B1
SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1	SOX2
SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU	SYK
TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1
TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WHSC1L1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

Α	LK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	KIT	EGFR
Ε	TV4	ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KMT2A (MLL)
Λ	1SH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
R	ARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**	TMPRSS2

^{*}TERC is an ncRNA

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER GENOMIC SIGNATURES

Microsatellite (MS) status Tumor Mutational Burden (TMB)

^{**}Promoter region of TERT is interrogated



APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.

ABOUT FOUNDATIONONE CDX

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing

Please refer to technical information for performance specification details: www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI) and tumor mutational burden (TMB) using DNA isolated from formalin-fixed paraffin embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLE

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes. Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X).

Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g.,gene fusions). Additionally, genomic signatures including microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: The association of a therapy with a genomic alteration or signature does not necessarily indicate pharmacologic effectiveness (or lack thereof); no association of a therapy with a genomic alteration or signature does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness).

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Alterations and Therapies Genomic Signatures

Appear at the top of the report, but are not ranked higher than Gene Alterations.

Gene Alterations

Therapies approved in the EU (In Patient's Tumor Type) → Therapies approved in the EU (In Other Tumor Type) → Clinical Trial Options → No Known Options (If multiple findings exist within any of these categories, the results are listed alphabetically by gene name.)

Therapies

Sensitizing therapies → Resistant therapies. (If multiple therapies exist within any of these categories, they are listed alphabetically by therapy name.)

Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

Limitations

- 1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established.
- 2. TMB by F1CDx is defined based on counting the total number of all synonymous and nonsynonymous variants present at 5% allele frequency or greater (after filtering) and reported as mutations per megabase (mut/Mb) unit rounded to the nearest integer. The clinical validity of TMB defined by this panel has not been established.

APPENDIX

About FoundationOne®CDx

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
os	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
ткі	Tyrosine kinase inhibitor

The median exon coverage for this sample is 733X

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- Gatalica Z, Snyder C, Maney T, et al. (2014) Programmed cell death 1 (PD-1) and its ligand (PD-L1) in common cancers and their correlation with molecular cancer type. Cancer Epidemiol Biomarkers Prev 23(12):2965-70
- Kroemer G, Galluzzi L, Zitvogel L, et al. (2015) Colorectal cancer: the first neoplasia found to be under immunosurveillance and the last one to respond to immunotherapy? Oncoimmunology 4(7):e1058597
- Lal N, Beggs AD, Willcox BE, et al. (2015) An immunogenomic stratification of colorectal cancer: Implications for development of targeted immunotherapy. Oncoimmunology 4(3):e976052
- 4. Overman et al., 2016; ASCO Abstract 3501
- Le DT, Uram JN, Wang H, et al. (2015) PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N Engl J Med ePub May 2015
- 6. ASCO-SITC 2016; Abstract P60
- Warth A, Körner S, Penzel R, et al. (2016) Microsatellite instability in pulmonary adenocarcinomas: a comprehensive study of 480 cases. Virchows Arch 468(3):313-9
- Ninomiya H, Nomura K, Satoh Y, et al. (2006) Genetic instability in lung cancer: concurrent analysis of chromosomal, mini- and microsatellite instability and loss of heterozygosity. Br J Cancer 94(10):1485-91
- Woenckhaus M, Stoehr R, Dietmaier W, et al. (2003)
 Microsatellite instability at chromosome 8p in nonsmall cell
 lung cancer is associated with lymph node metastasis and
 souamous differentiation. Int J Oncol 23(5):1357-63
- Chang JW, Chen YC, Chen CY, et al. (2000) Correlation of genetic instability with mismatch repair protein expression and p53 mutations in nonsmall cell lung cancer. Clin Cancer Res 6(5):1639-46
- Fong KM, Zimmerman PV, Smith PJ (1995) Microsatellite instability and other molecular abnormalities in non-small cell lung cancer. Cancer Res 55(1):28-30
- Hansen LT, Thykjaer T, Ørntoft TF, et al. (2003) The role of mismatch repair in small-cell lung cancer cells. Eur J Cancer 39(10):1456-67
- Kocarnik JM, Shiovitz S, Phipps AI (2015) Molecular phenotypes of colorectal cancer and potential clinical applications. Gastroenterol Rep (Oxf) 3(4):269-76
- You JF, Buhard O, Ligtenberg MJ, et al. (2010) Tumours with loss of MSH6 expression are MSI-H when screened with a pentaplex of five mononucleotide repeats. Br J Cancer 103(12):1840-5
- 15. Bairwa NK, Saha A, Gochhait S, et al. (2014) Microsatellite instability: an indirect assay to detect defects in the cellular mismatch repair machinery. Methods Mol Biol 1105:497-509
- 16. Boland CR, Thibodeau SN, Hamilton SR, et al. (1998) A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. Cancer Res 58(22):5248-57
- Pawlik TM, Raut CP, Rodriguez-Bigas MA (2004) Colorectal carcinogenesis: MSI-H versus MSI-L. Dis Markers 20(4-5): 199-206
- Boland CR, Goel A (2010) Microsatellite instability in colorectal cancer. Gastroenterology 138(6):2073-2087.e3
- Snyder A, Makarov V, Merghoub T, et al. (2014) Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med 371(23):2189-99
- 20. Rosenberg JE, Hoffman-Censits J, Powles T, et al. (2016) Atezolizumab in patients with locally advanced and metastatic urothelial carcinoma who have progressed following treatment with platinumbased chemotherapy: a single-arm, multicentre, phase 2 trial. Lancet 387(10031):1909-20
- 21. Johnson DB, Frampton GM, Rioth MJ, et al. (2016) Targeted next generation sequencing identifies markers of response to PD-1 blockade. Cancer Immunol Res ePub Sep 2016
- 22. Balar AV, Galsky MD, Rosenberg JE, et al. (2017) Atezolizumab as first-line treatment in cisplatinineligible patients with locally advanced and metastatic urothelial carcinoma: a single-arm, multicentre, phase 2 trial. Lancet 389(10064):67-76

- 23. Miao D, Margolis CA, Vokes NI, et al. (2018) Genomic correlates of response to immune checkpoint blockade in microsatellitestable solid tumors. Nat Genet 50(9):1271-1281
- 24. Dong ZY, Zhong WZ, Zhang XC, et al. (2016) Potential Predictive Value of TP53 and KRAS Mutation Status for Response to PD-1 Blockade Immunotherapy in Lung Adenocarcinoma. Clin Cancer Res
- Rizvi NA, Hellmann MD, Snyder A, et al. (2015) Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science 348(6230):124-8
- Mehnert JM, Panda A, Zhong H, et al. (2016) Immune activation and response to pembrolizumab in POLEmutant endometrial cancer. J Clin Invest 126(6):2334-40
- 27. Santin AD, Bellone S, Buza N, et al. (2016) Regression of chemotherapy-resistant Polymerase epsilon (POLE) ultramutated and MSH6 hyper-mutated endometrial tumors with nivolumab. Clin Cancer Res ePub Aug 2016
- Johanns TM, Miller CA, Dorward IG, et al. (2016) Immunogenomics of Hypermutated Glioblastoma: a Patient with Germline POLE Deficiency Treated with Checkpoint Blockade Immunotherapy. Cancer Discov ePub Sep 2016
- 29. Bouffet E, Larouche V, Campbell BB, et al. (2016) Immune Checkpoint Inhibition for Hypermutant Glioblastoma Multiforme Resulting From Germline Biallelic Mismatch Repair Deficiency. J Clin Oncol ePub Mar 2016
- Fabrizio DA, George TJ, Dunne RF, et al. (2018) Beyond microsatellite testing: assessment of tumor mutational burden identifies subsets of colorectal cancer who may respond to immune checkpoint inhibition. J Gastrointest Oncol 9(4): 610-617
- 31. Van Allen EM, Miao D, Schilling B, et al. (2015) Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. Science 350(6257):207-11
- 32. Legrand et al., 2018; ASCO Abstract 12000
- **33.** Spigel et al., 2016; ASCO Abstract 9017
- 34. Schwartz et al., 2016; ASCO Abstract 8533
- Xiao D, Pan H, Li F, et al. (2016) Analysis of ultra-deep targeted sequencing reveals mutation burden is associated with gender and clinical outcome in lung adenocarcinoma. Oncotarget 7(16):22857-64
- 36. Shim HS, Kenudson M, Zheng Z, et al. (2015) Unique Genetic and Survival Characteristics of Invasive Mucinous Adenocarcinoma of the Lung. J Thorac Oncol 10(8):1156-62
- Govindan R, Ding L, Griffith M, et al. (2012) Genomic landscape of non-small cell lung cancer in smokers and never-smokers. Cell 150(6):1121-34
- **38.** Ding L, Getz G, Wheeler DA, et al. (2008) Somatic mutations affect key pathways in lung adenocarcinoma. Nature 455(7216):1069-75
- Imielinski M, Berger AH, Hammerman PS, et al. (2012) Mapping the hallmarks of lung adenocarcinoma with massively parallel sequencing. Cell 150(6):1107-20
- 40. Kim Y, Hammerman PS, Kim J, et al. (2014) Integrative and comparative genomic analysis of lung squamous cell carcinomas in East Asian patients. J Clin Oncol 32(2):121-8
- **41.** Pfeifer GP, You YH, Besaratinia A (2005) Mutations induced by ultraviolet light. Mutat Res 571(1-2):19-31
- Hill VK, Gartner JJ, Samuels Y, et al. (2013) The genetics of melanoma: recent advances. Annu Rev Genomics Hum Genet 14:257-79
- 43. Pfeifer GP, Denissenko MF, Olivier M, et al. (2002) Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. Oncogene 21(48):7435-51
- 44. Cancer Genome Atlas Research Network, Kandoth C, Schultz N, et al. (2013) Integrated genomic characterization of endometrial carcinoma. Nature 497(7447):67-73
- **45.** Briggs S, Tomlinson I (2013) Germline and somatic polymerase ε and δ mutations define a new class of hypermutated colorectal and endometrial cancers. J Pathol 230(2):148-53

- **46.** Heitzer E, Tomlinson I (2014) Replicative DNA polymerase mutations in cancer. Curr Opin Genet Dev 24:107-13
- 47. Cancer Genome Atlas Network (2012) Comprehensive molecular characterization of human colon and rectal cancer. Nature 487(7407):330-7
- Roberts SA, Gordenin DA (2014) Hypermutation in human cancer genomes: footprints and mechanisms. Nat Rev Cancer 14(12):786-800
- 49. Camidge et al., 2011; ASCO Abstract 2501
- 50. Bang et al., 2010; ASCO Abstract 3
- 51. Gandhi et al., 2015; ASCO Abstract 8019
- 52. Kwak EL, Bang YJ, Camidge DR, et al. (2010) Anaplastic lymphoma kinase inhibition in non-smallcell lung cancer. N Engl J Med 363(18):1693-703
- 53. Gettinger SN, Bazhenova LA, Langer CJ, et al. (2016) Activity and safety of brigatinib in ALK-rearranged non-small-cell lung cancer and other malignancies: a single-arm, open-label, phase I/2 trial. Lancet Oncol 17(12):1683-1696
- 54. Shaw AT, Kim DW, Nakagawa K, et al. (2013) Crizotinib versus chemotherapy in advanced ALKpositive lung cancer. N Engl J Med 368(25):2385-94
- 55. Shaw AT, Kim DW, Mehra R, et al. (2014) Ceritinib in ALK-rearranged non-small-cell lung cancer. N Engl J Med 370(13): 1189-97
- 56. Ou SI, Ahn JS, De Petris L, et al. (2015) Alectinib in Crizotinib-Refractory ALK-Rearranged Non-Small-Cell Lung Cancer: A Phase II Global Study. J Clin Oncol ePub Nov 2015
- 57. Solomon BJ, Kim DW, Wu YL, et al. (2018) Final Overall Survival Analysis From a Study Comparing First-Line Crizotinib Versus Chemotherapy in ALKMutation- Positive Non-Small-Cell Lung Cancer. J Clin Oncol 36(22):2251-2258
- 58. Mossé YP, Lim MS, Voss SD, et al. (2013) Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: a Children's Oncology Group phase 1 consortium study. Lancet Oncol 14(6):472-80
- 59. Mosse et al., 2012; ASCO 9500
- Chand D, Yamazaki Y, Ruuth K, et al. (2013) Cell culture and Drosophila model systems define three classes of anaplastic lymphoma kinase mutations in neuroblastoma. Dis Model Mech 6(2):373-82
- 61. Schönherr C, Ruuth K, Yamazaki Y, et al. (2011) Activating ALK mutations found in neuroblastoma are inhibited by Crizotinib and NVP-TAE684. Biochem J 440(3):405-13
- 62. Kim DW, Mehra R, Tan DS, et al. (2016) Activity and safety of ceritinib in patients with ALK-rearranged non-small-cell lung cancer (ASCEND-1): updated results from the multicentre, open-label, phase 1 trial. Lancet Oncol ePub Mar 2016
- 63. Camidge DR, Kim DW, Tiseo M, et al. (2018) Exploratory Analysis of Brigatinib Activity in Patients With Anaplastic Lymphoma Kinase-Positive Non-Small-Cell Lung Cancer and Brain Metastases in Two Clinical Trials. J Clin Oncol: JCO2017775841
- 64. Huber et al., 2018; ASCO Abstract 9061
- **65.** Kim et al., 2018; ASCO Abstract 9009
- 66. Besse et al., 2018; ASCO Abstract 9032
- 67. Shaw et al., 2018; AACR Abstract CT044
- 68. Yoda S, Lin JJ, Lawrence MS, et al. (2018) Sequential ALK Inhibitors Can Select for Lorlatinib-Resistant Compound ALK Mutations in ALK-Positive Lung Cancer. Cancer Discov 8(6): 714-779
- 69. Dagogo-Jack I, Brannon AR, Ferris LA, et al. (2018) Tracking the Evolution of Resistance to ALK Tyrosine Kinase Inhibitors through Longitudinal Analysis of Circulating Tumor DNA. JCO Precis Oncol 2018
- Hochmair MJ, Schwab S, Prosch H (2017) Complete remission of intrathecal metastases with lorlatinib therapy in a heavily pretreated ALK-positive lung cancer patient. Anticancer Drugs 28(8):928-930

- 71. Yuan C, Ma MJ, Parker JV, et al. (2017) Metastatic Anapestic Lymphoma Kinase-I (ALK-I)-Rearranged Inflammatory Myofibroblastic Sarcoma to the Brain with Leptomeningeal Involvement: Favorable Response to Serial ALK Inhibitors: A Case Report. Am J Case Rep 18:799-804
- 72. Shaw et al., 2018: ASCO Abstract 9008
- Horn L, Infante JR, Reckamp KL, et al. (2018) Ensartinib (X-396) in ALK-Positive Non-Small Cell Lung Cancer: Results from a Firstin-Human Phase I/II, Multicenter Study. Clin Cancer Res 24(12): 2771-2770
- 74. Drilon A, Siena S, Ou SI, et al. (2017) Safety and Antitumor Activity of the Multi-Targeted Pan-TRK, ROS1, and ALK Inhibitor Entrectinib (RXDX-101): Combined Results from Two Phase 1 Trials (ALKA-372-001 and STARTRK-1). Cancer Discov ePub Feb 2017
- Socinski MA, Goldman J, El-Hariry I, et al. (2013) A multicenter phase II study of ganetespib monotherapy in patients with genotypically defined advanced non-small cell lung cancer. Clin Cancer Res 19(11):3068-77
- 76. Koivunen JP, Mermel C, Zejnullahu K, et al. (2008) EML4-ALK fusion gene and efficacy of an ALK kinase inhibitor in lung cancer. Clin Cancer Res 14(13):4275-83
- Inamura K, Takeuchi K, Togashi Y, et al. (2009) EML4-ALK lung cancers are characterized by rare other mutations, a TTF-1 cell lineage, an acinar histology, and young onset. Mod Pathol 22(4): 508-15
- Shaw AT, Yeap BY, Mino-Kenudson M, et al. (2009) Clinical features and outcome of patients with nonsmall-cell lung cancer who harbor EML4-ALK. J Clin Oncol 27(26):4247-53
- Takahashi T, Sonobe M, Kobayashi M, et al. (2010) Clinicopathologic features of non-small-cell lung cancer with EML4-ALK fusion gene. Ann Surg Oncol 17(3):889-97
- Li Y, Li Y, Yang T, et al. (2013) Clinical significance of EML4-ALK fusion gene and association with EGFR and KRAS gene mutations in 208 Chinese patients with non-small cell lung cancer. PLoS ONE 8(1):e52093
- Li H, Pan Y, Li Y, et al. (2013) Frequency of well identified oncogenic driver mutations in lung adenocarcinoma of smokers varies with histological subtypes and graduated smoking dose. Lung Cancer 79(1):8-13
- Zhou JX, Yang H, Deng Q, et al. (2013) Oncogenic driver mutations in patients with non-small-cell lung cancer at various clinical stages. Ann Oncol 24(5):1319-25
- 83. 83 Takeuchi K, Choi YL, Togashi Y, et al. (2009) KIF5BALK, a novel fusion oncokinase identified by an immunohistochemistry-based diagnostic system for ALKpositive lung cancer. Clin Cancer Res 15(9):3143-9
- 84. To KF, Tong JH, Yeung KS, et al. (2013) Detection of ALK rearrangement by immunohistochemistry in lung adenocarcinoma and the identification of a novel EML4-ALK variant. J Thorac Oncol 8(7):883-91
- 85. Grande E, Bolós MV, Arriola E (2011) Targeting oncogenic ALK: a promising strategy for cancer treatment. Mol Cancer Ther 10(4): 569-79
- **86.** Peters S, Taron M, Bubendorf L, et al. (2013) Treatment and detection of ALK-rearranged NSCLC. Lung Cancer 81(2):145-54
- 87. Li T, Maus MK, Desai SJ, et al. (2014) Large-scale screening and molecular characterization of EML4-ALK fusion variants in archival non-small-cell lung cancer tumor specimens using quantitative reverse transcription polymerase chain reaction assays. J Thorac Oncol 9(1):18-25
- Heuckmann JM, Balke-Want H, Malchers F, et al. (2012)
 Differential protein stability and ALK inhibitor sensitivity of EML4-ALK fusion variants. Clin Cancer
- 89. Lei YY, Yang JJ, Zhang XC, et al. (2015) Anaplastic Lymphoma Kinase Variants and the Percentage of ALK-Positive Tumor Cells and the Efficacy of Crizotinib in Advanced NSCLC. Clin Lung Cancer ePub Sep 2015
- Choi YL, Takeuchi K, Soda M, et al. (2008) Identification of novel isoforms of the EML4-ALK transforming gene in non-small cell lung cancer. Cancer Res 68(13):4971-6

- Sasaki T, Rodig SJ, Chirieac LR, et al. (2010) The biology and treatment of EML4-ALK non-small cell lung cancer. Eur J Cancer 46(10):1773-80
- Takeuchi K, Choi YL, Soda M, et al. (2008) Multiplex reverse transcription-PCR screening for EML4-ALK fusion transcripts. Clin Cancer Res 14(20):6618-24
- 93. Zhang X, Zhang S, Yang X, et al. (2010) Fusion of EML4 and ALK is associated with development of lung adenocarcinomas lacking EGFR and KRAS mutations and is correlated with ALK expression. Mol Cancer 9:188
- 94. Maus MK, Stephens C, Zeger G, et al. (2012) Identification of Novel Variant of EML4-ALK Fusion Gene in NSCLC: Potential Benefits of the RT-PCR Method. Int J Biomed Sci 8(1):1-6
- 95. Sanders HR, Li HR, Bruey JM, et al. (2011) Exon scanning by reverse transcriptase-polymerase chain reaction for detection of known and novel EML4-ALK fusion variants in non-small cell lung cancer. Cancer Genet 204(1):45-52
- Soda M, Choi YL, Enomoto M, et al. (2007) Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. Nature 448(7153):561-6
- Yoshida T, Oya Y, Tanaka K, et al. (2016) Differential Crizotinib Response Duration Among ALK Fusion Variants in ALK-Positive Non-Small-Cell Lung Cancer. J Clin Oncol ePub Jun 2016
- 98. Juric et al., 2016; ASCO Abstract 568
- 99. Peguero et al., 2016; ASCO Abstract 2528
- 100. Tolaney et al., 2016; SABCS P4-22-12
- 101. Morschhauser et al., 2014: ASH Abstract 3067
- 102. Flaherty KT, Lorusso PM, Demichele A, et al. (2012) Phase I, dose-escalation trial of the oral cyclindependent kinase 4/6 inhibitor PD 0332991, administered using a 21-day schedule in patients with advanced cancer. Clin Cancer Res 18(2):568-76
- 103. Finn RS, Crown JP, Lang I, et al. (2014) The cyclindependent kinase 4/6 inhibitor palbociclib in combination with letrozole versus letrozole alone as first-line treatment of oestrogen receptor-positive, HER2-negative, advanced breast cancer (PALOMA-1/TRIO-18): a randomized phase 2 study. Lancet Oncol ePub Dec 2014
- 104. Infante JR, Cassier PA, Gerecitano JF, et al. (2016) A Phase I Study of the Cyclin-Dependent Kinase 4/6 Inhibitor Ribociclib (LEE011) in Patients with Advanced Solid Tumors and Lymphomas. Clin Cancer Res 22(23):5696-5705
- 105. Patnaik A, Rosen LS, Tolaney SM, et al. (2016) Efficacy and Safety of Abemaciclib, an Inhibitor of CDK4 and CDK6, for Patients with Breast Cancer, Non-Small Cell Lung Cancer, and Other Solid Tumors. Cancer Discov 6(7):740-53
- 106. Leonard JP, LaCasce AS, Smith MR, et al. (2012) Selective CDK4/6 inhibition with tumor responses by PD0332991 in patients with mantle cell lymphoma. Blood 119(20):4597-607
- 107. Cancer Genome Atlas Research Network (2014) Comprehensive molecular profiling of lung adenocarcinoma. Nature 511(7511):543-50
- 108. Reissmann PT, Koga H, Figlin RA, et al. (1999) Amplification and overexpression of the cyclin D1 and epidermal growth factor receptor genes in nonsmall-cell lung cancer. Lung Cancer Study Group. J Cancer Res Clin Oncol 125(2):61-70
- 109. Marchetti A, Doglioni C, Barbareschi M, et al. (1998 Cyclin D1 and retinoblastoma susceptibility gene alterations in nonsmall cell lung cancer. Int J Cancer 75(2):187-92
- 110. Sun W, Song L, Ai T, et al. (2013) Prognostic value of MET, cyclin D1 and MET gene copy number in nonsmall cell lung cancer. J Biomed Res 27(3):220-30
- 111. Elsheikh S, Green AR, Aleskandarany MA, et al. (2008) CCND1 amplification and cyclin D1 expression in breast cancer and their relation with proteomic subgroups and patient outcome. Breast Cancer Res Treat 109(2):325-35
- **112.** Fu M, Wang C, Li Z, et al. (2004) Minireview: Cyclin D1: normal and abnormal functions. Endocrinology 145(12):5439-47
- 113. Takahashi-Yanaga F, Sasaguri T (2008) GSK-3beta regulates cyclin D1 expression: a new target for chemotherapy. Cell Signal 20(4):581-9

- 114. Guagnano V, Kauffmann A, Wöhrle S, et al. (2012) FGFR genetic alterations predict for sensitivity to NVP-BGJ398,a selective pan-FGFR inhibitor. Cancer Discov ePub Sep 2012
- 115. Hagel M, Miduturu C, Sheets M, et al. (2015) First Selective Small Molecule Inhibitor of FGFR4 for the Treatment of Hepatocellular Carcinomas with an Activated FGFR4 Signaling Pathway. Cancer Discoy 5(4):424-37
- 116. Kim et al., 2016; EORTC-NCI-AACR Symposium Abstract 105A
- 117. Chan et al., 2017; AACR Abstract CT106/24
- 118. Kaibori M, Sakai K, Ishizaki M, et al. (2016) Increased FGF19 copy number is frequently detected in hepatocellular carcinoma with a complete response after sorafenib treatment. Oncotarget ePub Jun 2016
- 119. Packer LM, Pollock PM (2015) Paralog-Specific Kinase Inhibition of FGFR4: Adding to the Arsenal of Anti-FGFR Agents. Cancer Discov 5(4):355-7
- 120. Sawey ET, Chanrion M, Cai C, et al. (2011) Identification of a therapeutic strategy targeting amplified FGF19 in liver cancer by Oncogenomic screening. Cancer Cell 19(3):347-58
- 121. Desnoyers LR, Pai R, Ferrando RE, et al. (2008) Targeting FGF19 inhibits tumor growth in colon cancer xenograft and FGF19 transgenic hepatocellular carcinoma models. Oncogene 27(1): 85-97
- 122. Miura S, Mitsuhashi N, Shimizu H, et al. (2012) Fibroblast growth factor 19 expression correlates with tumor progression and poorer prognosis of hepatocellular carcinoma. BMC Cancer 12:56
- 123. Feng S, Dakhova O, Creighton CJ, et al. (2013) Endocrine fibroblast growth factor FGF19 promotes prostate cancer progression. Cancer Res 73(8):2551-62
- 124. Nagamatsu H, Teishima J, Goto K, et al. (2015) FGF19 promotes progression of prostate cancer. Prostate ePub Apr 2015
- 125. Xie MH, Holcomb I, Deuel B, et al. (1999) FGF-19, a novel fibroblast growth factor with unique specificity for FGFR4. Cytokine 11(10):729-35
- **126.** Katoh M (2002) WNT and FGF gene clusters (review). Int J Oncol 21(6):1269-73
- 127. Kan Z, Zheng H, Liu X, et al. (2013) Whole-genome sequencing identifies recurrent mutations in hepatocellular carcinoma. Genome Res 23(9):1422-33
- 128. Tekin M, Hişmi BO, Fitoz S, et al. (2007) Homozygous mutations in fibroblast growth factor 3 are associated with a new form of syndromic deafness characterized by inner ear agenesis, microtia, and microdontia. Am J Hum Genet 80(2): 338-44
- 129. Arao T, Ueshima K, Matsumoto K, et al. (2013) FGF3/FGF4 amplification and multiple lung metastases in responders to sorafenib in hepatocellular carcinoma. Hepatology 57(4): 1407-15
- 130. Yamada T, Abei M, Danjoh I, et al. (2015) Identification of a unique hepatocellular carcinoma line, Li-7, with CD13(+) cancer stem cells hierarchy and population change upon its differentiation during culture and effects of sorafenib. BMC Cancer 15:260
- 131. Kratochwil K, Galceran J, Tontsch S, et al. (2002) FGF4, a direct target of LEF1 and Wnt signaling, can rescue the arrest of tooth organogenesis in Lef1(-/-) mice. Genes Dev 16(24):3173-85
- 132. Scherz PJ, Harfe BD, McMahon AP, et al. (2004) The limb bud Shh-Fgf feedback loop is terminated by expansion of former ZPA cells. Science 305(5682):396-9
- 133. Zaharieva BM, Simon R, Diener PA, et al. (2003) Highthroughput tissue microarray analysis of 11q13 gene amplification (CCND1, FGF3, FGF4, EMS1) in urinary bladder cancer. J Pathol 201(4):603-8
- 134. Arai H, Ueno T, Tangoku A, et al. (2003) Detection of amplified oncogenes by genome DNA microarrays in human primary esophageal squamous cell carcinoma: comparison with conventional comparative genomic hybridization analysis. Cancer Genet Cytogenet 146(1):16-21

- 135. Ribeiro IP, Marques F, Caramelo F, et al. (2014) Genetic imbalances detected by multiplex ligationdependent probe amplification in a cohort of patients with oral squamous cell carcinoma-the first step towards clinical personalized medicine. Tumour Biol 35(5):4687-95
- 136. Schulze K, Imbeaud S, Letouzé E, et al. (2015) Exome sequencing of hepatocellular carcinomas identifies new mutational signatures and potential therapeutic targets. Nat Genet 47(5): 505-11
- 137. Bivona TG, Hieronymus H, Parker J, et al. (2011) FAS and NF-κB signalling modulate dependence of lung cancers on mutant EGFR. Nature 471(7339):523-6
- 138. Giannikopoulos et al., 2014; ASCO Abstract 8083
- 139. Zhao Z, Zhong X, Wu T, et al. (2014) Identification of a NFKBIA polymorphism associated with lower NFKBIA protein levels and poor survival outcomes in patients with glioblastoma multiforme. Int J Mol Med 34(5):1233-40
- 140. Geng P, Ou J, Li J, et al. (2015) Genetic Association Between NFKBIA -881A>G Polymorphism and Cancer Susceptibility. Medicine (Baltimore) 94(31):e1024
- 141. Zhang M, Huang J, Tan X, et al. (2015) Common Polymorphisms in the NFKBIA Gene and Cancer Susceptibility: A Meta-Analysis. Med Sci Monit 21:3186-96
- **142.** Weniger MA, Küppers R (2016) NF-κB deregulation in Hodgkin lymphoma. Semin Cancer Biol ePub May 2016
- 143. Liu X, Yu H, Yang W, et al. (2010) Mutations of NFKBIA in biopsy specimens from Hodgkin lymphoma. Cancer Genet Cytogenet 197(2):152-7
- 144. Lake A, Shield LA, Cordano P, et al. (2009) Mutations of NFKBIA, encoding IkappaB alpha, are a recurrent finding in classical Hodgkin lymphoma but are not a unifying feature of non-EBVassociated cases. Int J Cancer 125(6):1334-42
- 145. Birnstiel ML, Jacob J, Sirlin JL (1965) Analysis of nucleolar RNA synthesis in dipteran salivary glands. Arch Biol (Liege) 76(2): 565-89
- 146. Cabannes E, Khan G, Aillet F, et al. (1999) Mutations in the IkBa gene in Hodgkin's disease suggest a tumour suppressor role for IkappaBalpha. Oncogene 18(20):3063-70
- 147. Bredel M, Scholtens DM, Yadav AK, et al. (2011) NFKBIA deletion in glioblastomas. N Engl J Med 364(7):627-37
- 148. Patanè M, Porrati P, Bottega E, et al. (2013) Frequency of NFKBIA deletions is low in glioblastomas and skewed in glioblastoma neurospheres. Mol Cancer 12:160
- 149. Gao J, Aksoy BA, Dogrusoz U, et al. (2013) Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6(269):pl1
- 150. Zack TI, Schumacher SE, Carter SL, et al. (2013) Pancancer patterns of somatic copy number alteration. Nat Genet 45(10): 1134-1140
- 151. Beroukhim R, Mermel CH, Porter D, et al. (2010) The landscape of somatic copy-number alteration across human cancers. Nature 463(7283):899-905
- 152. Hsu DS, Acharya CR, Balakumaran BS, et al. (2009) Characterizing the developmental pathways TTF-1, NKX2-8, and PAX9 in lung cancer. Proc Natl Acad Sci USA 106(13):5312-7
- 153. Yang L, Lin M, Ruan WJ, et al. (2012) Nkx2-1: a novel tumor biomarker of lung cancer. J Zhejiang Univ Sci B 13(11):855-66
- 154. Cancer Genome Atlas Research Network (2012) Comprehensive genomic characterization of squamous cell lung cancers. Nature 489(7417):519-25
- 155. Kumar A, Coleman I, Morrissey C, et al. (2016) Substantial interindividual and limited intraindividual genomic diversity among tumors from men with metastatic prostate cancer. Nat Med 22(4):369-78
- 156. Landa I, Ibrahimpasic T, Boucai L, et al. (2016) Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. J Clin Invest ePub Feb 2016
- 157. Jiao Y, Yonescu R, Offerhaus GJ, et al. (2014) Wholeexome sequencing of pancreatic neoplasms with acinar differentiation. J Pathol 232(4):428-35

- 158. Jones S, Stransky N, McCord CL, et al. (2014) Genomic analyses of gynaecologic carcinosarcomas reveal frequent mutations in chromatin remodelling genes. Nat Commun 5:5006
- 159. Nakamura N, Miyagi E, Murata S, et al. (2002) Expression of thyroid transcription factor-1 in normal and neoplastic lung tissues. Mod Pathol 15(10):1058-67
- 160. Moldvay J, Jackel M, Bogos K, et al. (2004) The role of TTF-1 in differentiating primary and metastatic lung adenocarcinomas. Pathol Oncol Res 10(2):85-8
- 161. Gilbert-Sirieix M, Makoukji J, Kimura S, et al. (2011) Wnt/β-catenin signalling pathway is a direct enhancer of thyroid transcription factor-1 in human papillary thyroid carcinoma cells. PLoS ONE 6(7):e22280
- 162. Robens J, Goldstein L, Gown AM, et al. (2010) Thyroid transcription factor-1 expression in breast carcinomas. Am J Surg Pathol 34(12):1881-5
- 163. Ni YB, Tsang JY, Shao MM, et al. (2014) TTF-1 expression in breast carcinoma: an unusual but real phenomenon. Histopathology 64(4):504-11
- 164. Tsai LH, Chen PM, Cheng YW, et al. (2014) LKB1 loss by alteration of the NKX2-1/p53 pathway promotes tumor malignancy and predicts poor survival and relapse in lung adenocarcinomas. Oncogene 33(29):3851-60
- 165. Tan D, Li Q, Deeb G, et al. (2003) Thyroid transcription factor-1 expression prevalence and its clinical implications in nonsmall cell lung cancer: a high-throughput tissue microarray and immunohistochemistry study. Hum Pathol 34(6):597-604
- 166. Haque AK, Syed S, Lele SM, et al. (2002) Immunohistochemical study of thyroid transcription factor-1 and HER2/neu in nonsmall cell lung cancer: strong thyroid transcription factor-1 expression predicts better survival. Appl Immunohistochem Mol Morphol 10(2):103-9
- 167. Pelosi G, Fraggetta F, Pasini F, et al. (2001) Immunoreactivity for thyroid transcription factor-1 in stage I non-small cell carcinomas of the lung. Am J Surg Pathol 25(3):363-72
- 168. Yamaguchi T, Hosono Y, Yanagisawa K, et al. (2013) NKX2-I/ TTF-1: an enigmatic oncogene that functions as a doubleedged sword for cancer cell survival and progression. Cancer Cell 23(6):718-23
- 169. Mu D (2013) The complexity of thyroid transcription factor 1 with both pro- and anti-oncogenic activities. J Biol Chem 288(35):24992-5000
- 170. Hamdan H, Liu H, Li C, et al. (1998) Structure of the human Nkx2.1 gene. Biochim Biophys Acta 1396(3):336-48
- 171. Kwei KA, Kim YH, Girard L, et al. (2008) Genomic profiling identifies TITF1 as a lineage-specific oncogene amplified in lung cancer. Oncogene 27(25):3635-40
- 172. Hirai H, Arai T, Okada M, et al. (2010) MK-1775, a small molecule Weel inhibitor, enhances anti-tumor efficacy of various DNA-damaging agents, including 5-fluorouracil. Cancer Biol Ther 9(7):514-22
- 173. Bridges KA, Hirai H, Buser CA, et al. (2011) MK-1775, a novel Wee1 kinase inhibitor, radiosensitizes p53-defective human tumor cells. Clin Cancer Res 17(17):5638-48
- 174. Rajeshkumar NV, De Oliveira E, Ottenhof N, et al. (2011) MK-1775, a potent Wee1 inhibitor, synergizes with gemcitabine to achieve tumor regressions, selectively in p53-deficient pancreatic cancer xenografts. Clin Cancer Res 17(9):2799-806
- 175. Osman AA, Monroe MM, Ortega Alves MV, et al. (2015) Wee-1 kinase inhibition overcomes cisplatin resistance associated with high-risk TP53 mutations in head and neck cancer through mitotic arrest followed by senescence. Mol Cancer Ther 14(2):608-19
- 176. Xu L, Huang CC, Huang W, et al. (2002) Systemic tumortargeted gene delivery by anti-transferrin receptor scFvimmunoliposomes. Mol Cancer Ther 1(5):337-46
- 177. Xu L, Tang WH, Huang CC, et al. (2001) Systemic p53 gene therapy of cancer with immunolipoplexes targeted by antitransferrin receptor scFv. Mol Med 7(10):723-34
- 178 Camp ER, Wang C, Little EC, et al. (2013) Transferrin receptor targeting nanomedicine delivering wildtype p53 gene sensitizes pancreatic cancer to gemcitabine therapy. Cancer Gene Ther 20(4):222-8

- 179. Kim SS, Rait A, Kim E, et al. (2015) A tumor-targeting p53 nanodelivery system limits chemoresistance to temozolomide prolonging survival in a mouse model of glioblastoma multiforme. Nanomedicine 11(2):301-11
- 180. Pirollo KF, Nemunaitis J, Leung PK, et al. (2016) Safety and Efficacy in Advanced Solid Tumors of a Targeted Nanocomplex Carrying the p53 Gene Used in Combination with Docetaxel: A Phase 1b Study. Mol Ther 24(9):1697-706
- 181. Hajdenberg et al., 2012; ASCO Abstract e15010
- 182. Leijen S, van Geel RM, Pavlick AC, et al. (2016) Phase I Study Evaluating WEE1 Inhibitor AZD1775 As Monotherapy and in Combination With Gemcitabine, Cisplatin, or Carboplatin in Patients With Advanced Solid Tumors. J Clin Oncol ePub Sep 2016
- 183. Oza et al., 2015; ASCO Abstract 5506
- 184. Leijen et al., 2015; ASCO Abstract 2507
- 185. Ma CX, Cai S, Li S, et al. (2012) Targeting Chk1 in p53-deficient triple-negative breast cancer is therapeutically beneficial in human-in-mouse tumor models. J Clin Invest 122(4):1541-52
- **186.** Mogi A, Kuwano H (2011) TP53 mutations in nonsmall cell lung cancer. J Biomed Biotechnol 2011:583929
- 187. Tekpli X, Landvik NE, Skaug V, et al. (2013) Functional effect of polymorphisms in 15q25 locus on CHRNA5 mRNA, bulky DNA adducts and TP53 mutations. Int J Cancer 132(8):1811-20
- 188. Vignot S, Frampton GM, Soria JC, et al. (2013) Next generation sequencing reveals high concordance of recurrent somatic alterations between primary tumor and metastases from patients with non-smallcell lung cancer. J Clin Oncol 31(17): 2167-72
- 189. Maeng CH, Lee HY, Kim YW, et al. (2013) Highthroughput molecular genotyping for small biopsy samples in advanced non-small cell lung cancer patients. Anticancer Res 33(11): 5127-33
- 190. Cortot AB, Younes M, Martel-Planche G, et al. (2014) Mutation of TP53 and alteration of p14(arf) expression in EGFR- and KRAS-mutated lung adenocarcinomas. Clin Lung Cancer 15(2): 124-30
- 191. Itakura M, Terashima Y, Shingyoji M, et al. (2013) High CC chemokine receptor 7 expression improves postoperative prognosis of lung adenocarcinoma patients. Br J Cancer 109(5):1100-8
- 192. Seo JS, Ju YS, Lee WC, et al. (2012) The transcriptional landscape and mutational profile of lung adenocarcinoma. Genome Res 22(11):2109-19
- 193. Brown CJ, Lain S, Verma CS, et al. (2009) Awakening guardian angels: drugging the p53 pathway. Nat Rev Cancer 9(12): 862-73
- 194. Joerger AC, Fersht AR (2008) Structural biology of the tumor suppressor p53. Annu Rev Biochem 77:557-82
- 195. Kato S, Han SY, Liu W, et al. (2003) Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. Proc Natl Acad Sci USA 100(14):8424-9
- 196. Kamada R, Nomura T, Anderson CW, et al. (2011) Cancerassociated p53 tetramerization domain mutants: quantitative analysis reveals a low threshold for tumor suppressor inactivation. J Biol Chem 286(1):252-8
- 197. Bougeard G, Renaux-Petel M, Flaman JM, et al. (2015) Revisiting Li-Fraumeni Syndrome From TP53 Mutation Carriers. J Clin Oncol 33(21):2345-52
- 198. Sorrell AD, Espenschied CR, Culver JO, et al. (2013) Tumor protein p53 (TP53) testing and Li-Fraumeni syndrome: current status of clinical applications and future directions. Mol Diagn Ther 17(1):31-4
- 199. Nichols KE, Malkin D, Garber JE, et al. (2001) Germline p53 mutations predispose to a wide spectrum of early-onset cancers. Cancer Epidemiol Biomarkers Prev 10(2):83-7
- **200.** Taubert H, Meye A, Würl P (1998) Soft tissue sarcomas and p53 mutations. Mol Med 4(6):365-72

- 201. Kleihues P, Schäuble B, zur Hausen A, et al. (1997) Tumors associated with p53 germline mutations: a synopsis of 91 families. Am J Pathol 150(1):1-13
- 202. Gonzalez KD, Noltner KA, Buzin CH, et al. (2009) Beyond Li Fraumeni Syndrome: clinical characteristics of families with p53 germline mutations. J Clin Oncol 27(8):1250-6
- 203. Lalloo F, Varley J, Ellis D, et al. (2003) Prediction of pathogenic mutations in patients with early-onset breast cancer by family history. Lancet 361(9363):1101-2
- 204. Ohe et al., 2015; ASCO Abstract 8061
- 205. Seto T, Kiura K, Nishio M, et al. (2013) CH5424802 (R05424802) for patients with ALK-rearranged advanced non-small-cell lung cancer (AF-001JP study): a single-arm, open-label, phase 1-2 study. Lancet Oncol 14(7):590-8
- 206. Gadgeel SM, Gandhi L, Riely GJ, et al. (2014) Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant ALK-rearranged non-smallcell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. Lancet Oncol 15(10):1119-28
- 207. Peters S, Camidge DR, Shaw AT, et al. (2017) Alectinib versus Crizotinib in Untreated ALK-Positive Non-Small-Cell Lung Cancer. N Engl J Med 377(9):829-838
- 208. Hida T, Nokihara H, Kondo M, et al. (2017) Alectinib versus crizotinib in patients with ALK-positive nonsmall-cell lung cancer (J-ALEX): an open-label, randomised phase 3 trial. Lancet 390(10089):29-39
- 209. Novello S, Mazières J, Oh IJ, et al. (2018) Alectinib versus chemotherapy in crizotinib-pretreated anaplastic lymphoma kinase (ALK)-positive nonsmall-cell lung cancer: results from the phase III ALUR study. Ann Oncol 29(6):1409-1416
- 210. Shaw AT, Gandhi L, Gadgeel S, et al. (2015) Alectinib in ALK-positive, crizotinib-resistant, non-small-cell lung cancer: a single-group, multicentre, phase 2 trial. Lancet Oncol ePub Dec 2015
- **211.** Lin et al., 2018; ASCO Abstract 9093
- 212. Gainor JF, Sherman CA, Willoughby K, et al. (2015) Alectinib salvages CN5 relapses in ALK-positive lung cancer patients previously treated with crizotinib and ceritinib. J Thorac Oncol 10(2):232-6
- Ajimizu H, Kim YH, Mishima M (2015) Rapid response of brain metastases to alectinib in a patient with non-small-cell lung cancer resistant to crizotinib. Med Oncol 32(2):477
- 214. Ou SH, Sommers KR, Azada MC, et al. (2015) Alectinib induces a durable (>15 months) complete response in an ALK-positive non-small cell lung cancer patient who progressed on crizotinib with diffuse leptomeningeal carcinomatosis. Oncologist 20(2): 224-6
- 215. Klempner SJ, Ou SH (2015) Anaplastic lymphoma kinase inhibitors in brain metastases from ALK+ nonsmall cell lung cancer: hitting the target even in the CNS. Chin Clin Oncol 4(2): 20
- 216. Dempke WC, Edvardsen K, Lu S, et al. (2015) Brain Metastases in NSCLC - are TKIs Changing the Treatment Strategy? Anticancer Res 35(11):5797-806

- 217. Lin JJ, Kennedy E, Sequist LV, et al. (2016) Clinical Activity of Alectinib in Advanced RET-Rearranged Non-Small-Cell Lung Cancer. J Thorac Oncol ePub Aug 2016
- 218. Kim et al., 2016; ASCO Abstract 9007
- 219. Camidge et al., 2016; World Conference on Lung Cancer P3.02a-013
- 220. Zhang S, Anjum R, Squillace R, et al. (2016) The Potent ALK Inhibitor Brigatinib (AP26113) Overcomes Mechanisms of Resistance to First- and Second- Generation ALK Inhibitors in Preclinical Models. Clin Cancer Res
- 221. Siaw JT, Wan H, Pfeifer K, et al. (2016) Brigatinib, an anaplastic lymphoma kinase inhibitor, abrogates activity and growth in ALK-positive neuroblastoma cells, Drosophila and mice. Oncotarget 7(20):29011-22
- 222. Kim DW, Tiseo M, Ahn MJ, et al. (2017) Brigatinib in Patients With Crizotinib-Refractory Anaplastic Lymphoma Kinase-Positive Non-Small-Cell Lung Cancer: A Randomized, Multicenter Phase II Trial. J Clin Oncol: JCO2016715904
- 223. Felip et al., 2015; ASCO Abstract 8060
- 224. Soria JC, Tan DS, Chiari R, et al. (2017) First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, openlabel, phase 3 study. Lancet 389(10072):917-929
- 225. Felip et al., 2015; ELCC Abstract 141PD
- 226. Scagliotti et al., 2016; ESMO Abstract LBA42_PR
- 227. Crinò L, Ahn MJ, De Marinis F, et al. (2016) Multicenter Phase II Study of Whole-Body and Intracranial Activity With Ceritinib in Patients With ALK-Rearranged Non-Small-Cell Lung Cancer Previously Treated With Chemotherapy and Crizotinib: Results From ASCEND-2. J Clin Oncol 34(24):2866-73
- 228. Kodityal S, Elvin JA, Squillace R, et al. (2016) A novel acquired ALK F1245C mutation confers resistance to crizotinib in ALKpositive NSCLC but is sensitive to ceritinib. Lung Cancer 92:19-21
- 229. Katayama R, Friboulet L, Koike S, et al. (2014) Two novel ALK mutations mediate acquired resistance to the next-generation ALK inhibitor alectinib. Clin Cancer Res 20(22):5686-96
- 230. Ou SH, Greenbowe J, Khan ZU, et al. (2015) I1171 missense mutation (particularly I1171N) is a common resistance mutation in ALK-positive NSCLC patients who have progressive disease while on alectinib and is sensitive to ceritinib. Lung Cancer 88(2):231-4
- 231. Lu et al., 2016; ASCO Abstract 9058
- 232. Solomon BJ, Mok T, Kim DW, et al. (2014) First-line crizotinib versus chemotherapy in ALK-positive lung cancer. N Engl J Med 371(23):2167-77
- 233. Butrynski JE, D'Adamo DR, Hornick JL, et al. (2010) Crizotinib in ALK-rearranged inflammatory myofibroblastic tumor. N Engl J Med 363(18):1727-33
- 234. Pérot G, Soubeyran I, Ribeiro A, et al. (2014) Identification of a recurrent STRN/ALK fusion in thyroid carcinomas. PLoS ONE 9(1):e87170

- 235. Blackhall F, Kim DW, Besse B, et al. (2014) Patient reported outcomes and quality of life in PROFILE 1007: a randomized trial of crizotinib compared with chemotherapy in previously treated patients with ALK-positive advanced non-small-cell lung cancer. Thorac Oncol 9(11):1625-33
- 236. Shaw et al., 2016; ASCO Abstract 9066
- 237. Solomon BJ, Cappuzzo F, Felip E, et al. (2016) Intracranial Efficacy of Crizotinib Versus Chemotherapy in Patients With Advanced ALKPositive Non-Small-Cell Lung Cancer: Results From PROFILE 1014. J Clin Oncol 34(24):2858-65
- 238. Costa DB, Shaw AT, Ou SH, et al. (2015) Clinical Experience With Crizotinib in Patients With Advanced ALK-Rearranged Non-Small-Cell Lung Cancer and Brain Metastases. J Clin Oncol 33(77):1881-8
- 239. Johung KL, Yeh N, Desai NB, et al. (2016) Extended Survival and Prognostic Factors for Patients With ALK-Rearranged Non-Small-Cell Lung Cancer and Brain Metastasis. J Clin Oncol 34(2):123-9
- 240. Ou SH, Jänne PA, Bartlett CH, et al. (2014) Clinical benefit of continuing ALK inhibition with crizotinib beyond initial disease progression in patients with advanced ALK-positive NSCLC. Ann Oncol 25(2):415-22
- 241. Sledge GW, Toi M, Neven P, et al. (2017) MONARCH 2: Abemaciclib in Combination With Fulvestrant in Women With HR+/HER2-Advanced Breast Cancer Who Had Progressed While Receiving Endocrine Therapy. J Clin Oncol 35(25): 2875-2884
- 242. Dickler MN, Tolaney SM, Rugo HS, et al. (2017) MONARCH1, A Phase II Study of Abemaciclib, a CDK4 and CDK6 Inhibitor, as a Single Agent, in Patients with Refractory HR(+)/HER2(-) Metastatic Breast Cancer. Clin Cancer Res 23(17):5218-5224
- 243. Kim et al., 2015; ASCO Abstract 8047
- 244. Dickson MA, Tap WD, Keohan ML, et al. (2013) Phase II trial of the CDK4 inhibitor PD0332991 in patients with advanced CDK4-amplified well-differentiated or dedifferentiated liposarcoma. J Clin Oncol 31(16):2024-8
- 245. Turner NC, Ro J, André F, et al. (2015) Palbociclib in Hormone-Receptor-Positive Advanced Breast Cancer. N Engl J Med ePub Jun 2015
- 246. DeMichele A, Clark A, Tan KS, et al. (2014) CDK 4/6 Inhibitor Palbociclib (PD0332991) in Rb+ Advanced Breast Cancer: Phase II Activity, Safety and Predictive Biomarker Assessment. Clin Cancer Res ePub Dec 2014
- 247. Gopalan et al., 2014; ASCO Abstract 8077
- 248. Goldman et al., 2014; ASCO Abstract 8026
- 249. Yamada et al., 2015; AACR-NCI-EORTC Abstract B31
- 250. Geoerger B, Bourdeaut F, DuBois SG, et al. (2017) A Phase I Study of the CDK4/6 Inhibitor Ribociclib (LEE011) in Pediatric Patients with Malignant Rhabdoid Tumors, Neuroblastoma, and Other Solid Tumors. Clin Cancer Res